



G-Biosciences ♦ 1-800-628-7730 ♦ 1-314-991-6034 ♦ technical@GBiosciences.com

A Geno Technology, Inc. (USA) brand name

GET™ Viral Nucleic Acid-Mag

Genomic Efficient Technology for Viral Nucleic Acid
purification with silica magnetic beads
(Cat. # 786-1737, 786-1738)



think proteins! think G-Biosciences www.GBiosciences.com

INTRODUCTION 3

ITEMS SUPPLIED 4

STORAGE CONDITIONS 5

ADDITIONAL ITEMS NEEDED..... 5

IMPORTANT INFORMATION 5

PREPARATION BEFORE USE 5

PROTOCOL..... 6

 SERUM, PLASMA OR CELL CULTURE SUPERNATANT VIRAL SAMPLES 6

 SWAB (VIRAL SAMPLE) FROM NOSE, THROAT OR OTHER TEST AREA 7

TROUBLESHOOTING 8

RELATED PRODUCTS..... 9

INTRODUCTION

Get™ Viral Nucleic Acid-Mag Kit belongs to our series of kits based on Genomic Efficient Technology (GET) for purification of nucleic acids from diverse sample.

GET™ is based on a highly efficient Genomic lysis buffer that liberates nucleic acid from cellular protein complexes, making nucleic acids free and available for purification in pure form. Free nucleic acids, DNA templates, are immobilized, in the presence of high concentration of chaotropic agents, on silica surface. Following the capture of nucleic acid on the silica membrane or beads, a series of washing steps removes interfering impurities. In the final step, pure DNA template is eluted in concentrated form with GET Elution Buffer.

The Get™ Viral Nucleic Acid- Mag is based upon the principle of binding of nucleic acids on silica beads with magnetic core in presence of chaotropic salts. Nucleic acids are bound to the silica magnetic beads under high concentrations of chaotropic salts and the impurities are removed during the wash steps and nucleic acid eluted with GET Elution Buffer (Fig:1). The DNA or RNA extraction is rapid with less than 15-30 minutes hands- on time.

Furthermore, kit is optimized to obtain high quality viral nucleic acid with minimum to no loss of nucleic acid. The purified nucleic acid fraction obtained is concentrated and is suitable for PCR, RT-PCR, qPCR and qRT-PCR studies. These PCR studies are used for virus detection, virus load, and virus genotyping.

Get™ Viral Nucleic Acid-Mag is available in 50 and 100 prep sizes with the maximum biological sample volume of 200 µl per prep.

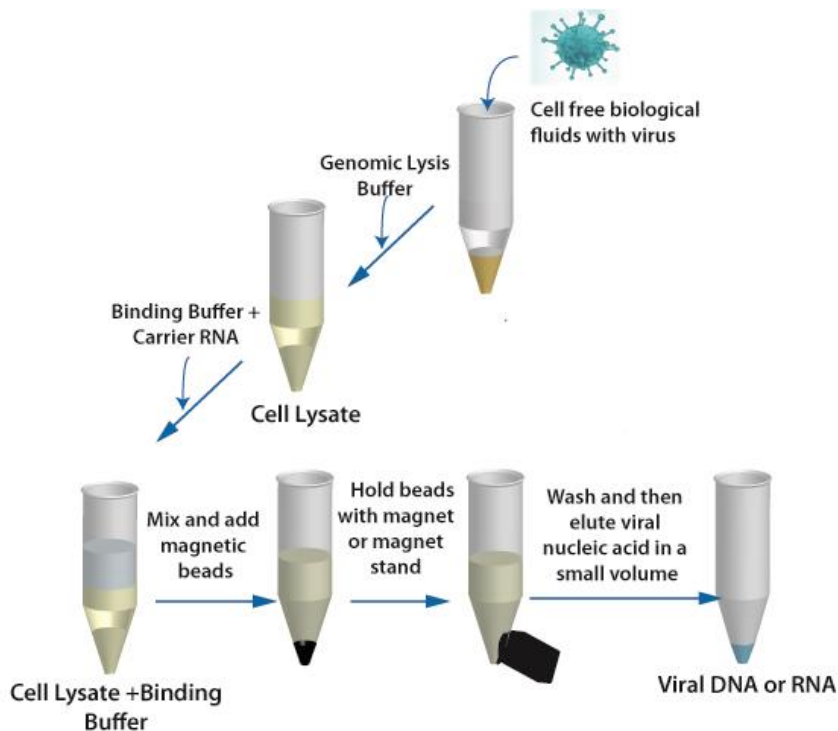


Fig:1

ITEMS SUPPLIED

Description	Cat. # 786-1737 50 preps	Cat. # 786-1738 100 preps
Genomic Lysis Buffer	30 ml	2 x 30 ml
GET Binding Buffer	50	2 x 50 ml
Poly A [Carrier RNA]	1 vial	2 vials
Longlife™ Proteinase K	0.5 ml	2 x 0.5 ml
GET Wash I	30 ml	2 x 30 ml
GET Wash II	20 ml	2x 20 ml
GET Elution Buffer	10 ml	10 ml
Silica Magnetic Beads	2.5 ml	2 x 2.5 ml

STORAGE CONDITIONS

The kit is supplied at ambient temperature. Upon receipt store the kit components as indicated on labels. If stored and used as directed, the kit is stable for 1 year.

ADDITIONAL ITEMS NEEDED

1. Magnetic stand (Cat. # 786-888) or a magnet
2. Absolute ethanol
3. 1.5 ml nuclease-free (sterile) microfuge tubes
4. (Optional) RNaseOUT™ (Cat. #786-70) for cleaning working areas and equipment.
5. Nuclease free tips for the pipettes.

IMPORTANT INFORMATION

- Get™ Viral Nucleic Acid is available in 50 prep (Cat. # 786-1737) and 100 prep (Cat. # 786-1738) sizes with sample volume of 200 µl. For more than 200µl sample increase the volume of magnetic beads and buffers added proportionally as per protocol.
- Since the sensitivity and titer of potential pathogen (virus) varies with samples, the end user needs take appropriate safety measures when handling.
- Sterile handling of reagents and sample should be carried out to avoid contamination of reagents and sample with bacteria or nuclease. Frequent change of gloves and cleaning of work area with RNaseOUT™ (Cat. #786-70) is recommended. Avoid touching the mouth of reagent bottles.
- For performing RT-PCR, PCR, qPCR or qRT-PCR experiments using viral nucleic acid, ensure that sample preparation, RT-PCR or PCR run, and electrophoresis are carried out in separate work areas to avoid cross-contamination.

PREPARATION BEFORE USE

1. Add 18 ml of molecular grade ethanol to 30 ml GET Wash I bottle and check the box on the bottle label to indicate ethanol has been added.
2. Add 80ml molecular grade ethanol to the GET Wash II bottle (20 ml) and check the box on the bottle label to indicate ethanol has been added.
3. Equilibrate GET Elution Buffer to 60°C.
4. Briefly centrifuge the Poly A [Carrier RNA] tube. Add 250 µl of GET Elution Buffer to 1 vial of Poly A [Carrier RNA]. Gently mix with pipette to dissolve. Aliquot 50µl per microfuge vial and store the vials at -20°C as stock solution.
5. Store Longlife™ Proteinase K in small aliquots at -20°C for long term use. Before use, Invert the Longlife™ Proteinase K tube 3-4 times to mix the enzyme suspension, then remove an aliquot for use.
6. **Working solution of GET Binding Buffer** (*Prepare fresh and do not store after use*): Thaw one vial of Poly A [Carrier RNA] (50 µl). Transfer the Poly A [Carrier RNA] solution to 10 ml of GET Binding Buffer. Briefly vortex to mix.

PROTOCOL

Serum, plasma or cell culture supernatant viral samples

1. Add 200 μ l of Genomic Lysis Buffer to 200 μ l of viral sample. Vortex to mix.
NOTE: For sample volume <200 μ l add PBS or 0.9%NaCl to make up the volume to 200 μ l.
NOTE: For sample volume >200 μ l increase the volume of reagents added proportionally.
2. Add 10 μ l Longlife™ Proteinase K suspension into the sample, mix and incubate at 55°C-60°C for 1 hr.
NOTE: Before use, Invert the Longlife™ Proteinase K tube 3-4 times to mix the enzyme suspension, then remove an aliquot for use.
NOTE: Do not exceed 60°C.
3. Add 800 μ l of working solution of GET Binding Buffer to the sample and vortex to mix
4. Add 50 μ l of Silica Magnetic Beads to sample tube and gently invert 4-5 times to mix. Incubate the sample for 5 minutes with gentle mixing on rotor at room temperature.
NOTE: Do not vortex or shake the sample containing silica magnetic beads vigorously.
5. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.
6. Gently remove and discard the supernatant without disturbing the beads.
7. Remove the tube from magnetic stand or magnet and add 0.5ml GET Wash I to the sample tube and invert the tube gently 4-5 times to mix.
8. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.
9. Gently remove and discard the supernatant without disturbing the beads.
10. Remove the tube from magnetic stand or magnet and add 0.5ml GET Wash II to the tube and invert the tube gently 4-5 times to mix.
11. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.
12. Gently remove and discard the supernatant without disturbing the beads.
13. Repeat step 10-12.
14. Ensure all the liquid is removed from the magnetic beads. Air dry the beads for 5-20 minutes.
15. Add 50 μ l of prewarmed GET Elution Buffer to the magnetic beads and resuspend the beads complex by brief vortex or shaking. Incubate for 15 minutes with gentle mixing on rotor at room temperature.
16. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube. Transfer the supernatant in a 1.5 ml nuclease free tube.
NOTE: Check the recovery. If recovery is poor, add 25-50 μ l prewarmed (50-60°C)

GET Elution Buffer to the magnetic beads and repeat steps 15-16. Combine with previous elution

17. Store the eluted DNA at 4°C for short term use or at -20°C for long term use. Eluted RNA should be stored at -80°C.

Swab (Viral sample) from nose, throat or other test area

1. Transfer swab containing specimen sample (nose, throat or other test area) in to a swab tube containing either PBS or 0.9 % NaCl. Vortex the tube for 1-2 minutes to release the specimen (virus) in the solution.

2. Remove the swab and centrifuge the tube at 15,000 g for 10 minutes at room temperature.

NOTE: *Alternatively, filtration can be used to remove cells from swabs.*

3. Transfer 200 µl supernatant into a nuclease free microfuge tube. Add 200 µl of Genomic Lysis Buffer, vortex to mix.

4. Add 10 µl Longlife™ Proteinase K suspension into the sample, mix and incubate at 55°C-60°C for 1 hr.

NOTE: *Before use, invert the Longlife™ Proteinase K tube 3-4 times to mix the enzyme suspension, then remove an aliquot for use.*

NOTE: *Do not exceed 60°C.*

5. Add 800 µl of working solution of GET Binding Buffer to the sample and vortex to mix.

6. Add 50 µl of Silica Magnetic Beads to sample tube and gently invert 4-5 times to mix. Incubate the sample for 5 minutes with gentle mixing on rotor at room temperature.

NOTE: *Do not vortex or shake the sample containing silica magnetic beads vigorously.*

7. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.

8. Gently remove and discard the supernatant without disturbing the beads.

9. Remove the tube from magnetic stand or magnet and add 0.5ml GET Wash I to the sample tube and invert the tube gently 4-5 times to mix.

10. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.

11. Gently remove and discard the supernatant without disturbing the beads.

12. Remove the tube from magnetic stand or magnet and add 0.5ml GET Wash II to the tube and invert the tube gently 4-5 times to mix.

13. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube.

14. Gently remove and discard the supernatant without disturbing the beads.

15. Repeat step 12-14.

16. Ensure all the liquid is removed from the magnetic beads. Air dry the beads for 5-20 minutes.

17. Add 50 μ l of prewarmed GET Elution Buffer to the magnetic beads and resuspend the beads complex by brief vortex or shaking. Incubate for 15 minutes with gentle mixing on rotor at room temperature.
18. Place the sample tube on magnetic stand or use magnet to immobilize the beads at one end of the tube. Transfer the supernatant in a 1.5 ml nuclease free tube.
NOTE: Check the recovery. If recovery is poor, add 25-50 μ l prewarmed (50-60°C) GET Elution Buffer to the magnetic beads and repeat steps 15-16. Combine with previous elution
19. Store the eluted DNA at 4°C for short term use or at -20°C for long term use. Eluted RNA should be stored at -80°C.

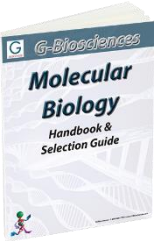
TROUBLESHOOTING

Issue	Suggested reason	Possible solution
Low nucleic acid yield or purity	Kit components are not stored properly	Store kit components as indicated in the label.
	Ethanol not added to the GET Wash I and II Buffers	Add absolute ethanol to GET Wash I and II Buffers before using
	Reagent and samples not properly mixed	Mix the sample tube well after addition of each reagent.
Low RNA yield	High levels of RNase activity	Create RNase-free work environment. Use RNase OUT to clean the working bench
		Process starting material immediately or store at -80°C until it is processed
		Use eluted RNA directly for downstream application or store at -80°C for later use
	Incomplete Proteinase K digestion	Thaw Longlife™ Proteinase K on ice and Resuspend Proteinase K solution with pipette before adding to the sample. Incubate for longer time if necessary.
Poor elution of nucleic acids	GET Elution Buffer provided in kit is not used	Use the GET Elution Buffer provided in the kit
		If using own water or GET Elution Buffer, ensure the pH

		of buffer is same as that of the GET Elution Buffer provided
--	--	--

RELATED PRODUCTS

Download our Molecular Biology Handbook.



<http://info2.gbiosciences.com/complete-molecular-biology-handbook>

For other related products, visit our website at www.GBiosciences.com or contact us.

Version1: 5/28/20



www.GBiosciences.com